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Preparation of various C-2 branched carbohydrates using intramolecular radical reactions

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Abstract

A new and efficient method for the facile synthesis of C-2 branched carbohydrates has been developed using an intramolecular radical cyclization-fragmentation reaction. The desired C-2 branched glucopyranosides were isolated in 40–84% yield. Additionally, an unexpected furanoside was obtained from a tributyltin iodide-promoted rearrangement of the radical intermediate. The C-2 formyl glycal was also isolated in good yield using tris(trimethylsily)silane (TTMSS) as the reducing agent. This method was extended to synthesize a β C-2 branched glucopyranoside, a C-2 branched galactoside and a C-2 cyano glucopyranoside. © 2000 Elsevier Science Ltd. All rights reserved.

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1. Introduction

While investigating the synthesis of the potential antiviral agents cyclophellitol 1 β and 1 α , an efficient method was required to construct α C-2 formyl glucopyranoside 4 α [1]. The sequence of reactions used in the synthesis of cyclophellitol was lengthy and repetitive, although the overall yield was acceptable [2]. The sequence required a Swern oxidation of the mannopyranoside 2, followed by a Wittig olefination to produce 3 in 70% yield (two steps). Hydroboration of the olefin 3 afforded a mixture of hydroxymethylglucopyranosides that was oxidized to give C-2 formyl adducts 4 $\alpha\beta$. The aldehydes were treated under mild basic conditions overnight to furnish the desired C-2 formyl glucopyranoside 4α (Scheme 1).

Various methods have been reported for the preparation of C-2 branched sugars [3]. Balasubramanian and co-workers published a short route to C-2 formyl glycals using the Vilsmeier–Haack reaction [4]. The reported reaction conditions, however, were too harsh for acid-labile protective groups. The most direct approach was accomplished by Kahne and co-workers using an intermolecular radical trapping of carbon monoxide (CO). However, the reported low yields (5-37%) and the required reaction conditions (1200-1400 psi) CO) made it less attractive as a general method for the preparation of C-2 branched sugars [5].

This paper describes a new method to synthesize α C-2 formyl carbohydrates using commercially available glucals by employing an intramolecular cyclization-fragmentation rad-

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Scheme 1. Synthesis of 2-*C*-formyl glucoside 3α. Reagents: (i) (a) DMSO, TFAA, TEA, (b) Wittig; (ii) (a) BH₃·DMS, THF, (b) DMSO, TFAA, TEA; (iii) TEA.

ical reaction (Scheme 2) [6]. The functionalizable formyl moiety is ideal to further branch the sugar if needed. The mechanism for the formation of the desired C-2 formyl sugar requires the generation of a secondary alkyl radical from iodoaldehydes 9 and 10. The resulting radical adds intramolecularly to the aldehyde group at the C-1 center of the sugar to give the highly reactive bicyclic alkoxy radical. The radical intermediate undergoes β scission to produce an α -isopropoxy radical, which upon reduction affords the formyltransfer products 11 and 12. A conceptually similar process was reported by Fraser-Reid and co-workers [7]. They reported a number of examples that transposes an aldehyde group upon β -scission onto the side chains of carbohydrates.

2. Results and discussion

2-Iodo glucopyranoside 7 was prepared in 75% yield from commercially available tri-O-acetyl-D-glucal (5) by addition of N-iodosuccinimide and commercially available dimethylvinyl carbinol (Scheme 2) [8]. Ozonolysis and reduction with methyl sulfide furnished the desired iodoaldehyde 9 in 96% yield. To test the feasibility of the radical cyclization, 7 was subjected to tributyltin hydride-promoted radical cyclization, which successfully furnished the cyclized product 13 in 68% yield as a 1:1 mixture of diastereomers.

The free-radical promoted cyclization of the aldehyde 9 produced a mixture of products in varying yields depending on the reaction conditions. The products from the radical reactions were separated using flash column chromatography. The desired formyl-transfer product 11 was isolated in 40% yield under optimal conditions (Table 1, Method A). In addition to the C-2 formyl glucopyranoside 11, the bicyclic alcohols $14\alpha\beta$ as a 1:1 mixture of diastereomers, the C-2 formyl glycal 17, and the directly reduced sugar 19 were isolated from the reaction. An unexpected furanoside 16 was isolated, and the structure was determined by careful spectroscopic analysis.



Scheme 2. Synthesis of the radical substrates and the products from the radical reactions. Reagents: (i) NIS, dimethylvinyl carbinol, acetonitrile; (ii) O₃, DMS; (iii) Bu₃SnH, AIBN, PhH.

Method	[9]°	Time (h) ^d	[11]	[14]	[16]	[17]	[19]
A	0.004	0.5			60		
A	0.005	2	40		27		23
A	0.01	0.5	16		42		
A	0.02	6	21	20	38	10	
A	0.03	5			42		9
В	0.006	5		17		65	
В	0.01	1.5	20	20		17	
В	0.021	3	30	13		32	
В	0.024	1.5	41	33		11	
В	0.024	3	17	13		31	

Representative reaction	n conditions and	l product distrib	ation for the	radical reaction a,b

T 11

^a Method A: typical reaction was conducted in benzene with 1.0–1.5 equiv of Bu₃SnH and 0.01–0.5 equiv of AIBN under reflux.

^b Method B: typical reaction was conducted in toluene with 1.0–1.5 equiv of TTMSS and 0.01–0.5 equiv of AIBN under reflux. ^c Initial concentrations (mol/L) of the deoxyiodoglucoside **9**.

d T

^d Time required to deliver the reducing agent using a syringe pump.



Scheme 3. Proposed mechanism for the formation of the furanoside and NOE data.

NMR analysis of 16 indicated the absence of large trans-diaxial coupling constants and axial-equatorial coupling constants typical for glucopyranosides. Instead, coupling constants expected from a furanoside ring system were observed. We propose that the furanoside 16 is formed from a tributyltin iodidecatalyzed rearrangement of the radical intermediate 14α (Scheme 3). The mechanism involves the complexation of the pyran oxygen with the iodostannane, followed by a pyranoside ring opening to give 16A. The resulting oxonium ion 16A would then be attacked by the C-4 acetate group to afford 16B, followed by an internal (or intermolecular) acetate transfer to give the furanoside 16. The stereochemistry of the alcohol was determined using NOE difference spectra (Scheme 3). The NOE difference spectra of H-3 (5.37 ppm) showed a strong enhancement of the signal for H-4 (4.74 ppm) and a weak enhancement of the signal for H-7 (3.86 ppm), H-2 (2.45 ppm) and OH (2.46 ppm). Irradiation of H-7 (3.86 ppm) showed weak enhancement of H-3 (5.37 ppm) but a strong enhancement of the signal for H-2 (2.45 ppm) and one of the methyl singlets (1.19 ppm). Irradiation of the methyl singlet at 1.19 ppm showed a strong NOE to both H-2 and H-7. which added further evidence that this methyl group is cis to both H-2 and H-7. Lastly, irradiation of the methyl singlet at 1.39 ppm showed a strong enhancement of OH (2.46 ppm) but a weak enhancement of H-7 and no NOE enhancement of H-2. The methyl singlet at 1.39 ppm was therefore assigned cis to OH and trans to both H-2 and H-7.

Chatgilialoglu and co-workers reported that tris(trimethylsilyl)silane (TTMSS) rivaled tri-

butyltin hydride in efficiency as a reducing agent with several advantages [9]. They found that the silane was less reactive as a hydrogen donor than the stannanes since the Si-H bond is 5 kcal mol⁻¹ stronger than the Sn-H bond. In addition, removal of the byproducts from the radical reaction using TTMSS was reported to be much easier than with the tin reagent. To diminish the formation of 14 and 19, the radical precursor 9 was subjected to TTMSS radical conditions. Furthermore, the sterically bulky nature of the intermediate tris(trimethylsilyl)silyl iodide was expected to function as a poor catalyst for the rearrangement needed to give the furanoside 16. As anticipated, the radical reactions using TTMSS (Table 1. Method B) did not form any of the furanoside 16. However, a 65% yield of the C-2 formyl glycal 17 was isolated. This route to C-2 formyl glycals is more general by allowing for acid-sensitive protecting groups than the known method via the Vilsmeier-Haack reaction. Along with the glycal, the bicyclic alcohols 14 $\alpha\beta$ were isolated in varying yields depending on the reaction conditions. The desired formyltransfer product 11 was isolated in 41% yield under optimal conditions. Efforts towards increasing the yield of 11 using in situ generation of the reducing agent and attempts using hexabutyldistannane under photolytic conditions were unsuccessful.

Table 2 Product distribution for the radical reaction

[10]	Time (h)	[12]	[15]	[18]	[21]
0.009 ^a	3.4	14	30	17	
0.009 ^a	6.5	20	20	21	
0.01 ^a	3	20	58		
0.02 ^a	6.5	11	60		
0.02 ^a	3	17	53		
0.004 ^ь	0.5				67
0.005 ^ь	3				82
0.03 ^ь	3				67
0.009°	3.5		45		

^a Typical reaction was conducted in benzene with 10a, 1.0–1.5 equiv of Bu₃SnH and 0.01–0.5 equiv of AIBN under reflux.

^b Typical reaction was conducted in benzene with **10b**, 1.0-1.5 equiv of Bu₃SnH and 0.01-0.5 equiv of AIBN under reflux.

^c Radical reaction with 10a using TTMSS as the reducing agent.

Conversion of the commercially available tri-O-benzyl-D-glucal (6) [8] by the same twostep procedure furnished the desired radical precursor 10 (Scheme 2). A solvent effect was observed on the iodoetherification of 6. Namely, a reaction using tetrahydrofuran as the solvent afforded the diaxial α glucoside 8α exclusively in 86% yield. However, using acetonitrile as the solvent, the diaxial glucoside 8α and the dieguatorial glucoside $\mathbf{8}\beta$ were isolated as a 4:1 mixture in 62% yield. The two diastereomers were separated and ozonized to give an 82% yield of the diaxial iodoaldehyde 10α and a 76% yield of the dieguatorial iodoaldehyde 10β . Easy access to the two isomers allowed us to investigate not only the α -formyl transfer reactions but also the feasibility of extending this process to generate the β -branched sugars.

The formyl-transfer reaction of 10α using tributyltin hydride gave 12 in 20% yield under optimal conditions (Table 2). Under most experimental conditions, the alcohols $15\alpha\beta$ were isolated as the major product. Formyl glycal 18 was isolated in 21% yield. No furanosides and only traces of the directly reduced product 20 were detected in the radical reactions of $10\alpha\beta$. Using TTMSS on the radical substrate 10α yielded the bicyclic alcohol 15 with only traces of other byproducts.

Extending the formyl-transfer reaction to β branched sugars, the iodoaldehyde **10** β was subjected to radical reaction conditions (Table 2). The reaction yielded the alcohol **21** as the major product with yields as high as 82% with no traces of the β C-2 formyl product.

The radical reactions for the tri-O-acetyl and the tri-O-benzyl glucal substrates furnished substantial amounts of the bicyclic alcohols 14, 15, and 21. Therefore, in order to increase the yields of 11 and 12, these intermediates were subjected to reaction conditions that regenerate the alkoxy radical, thus allowing the β -fission to complete the rearrangement process [10]. However, attempts at converting the alcohols to the desired formyl-transfer products were unsuccessful. In several cases the 1:1 mixture of the alcohols $14\alpha\beta$ epimerized to give mostly one isomer upon reaction with mercuric oxide and iodine under photolysis. This result indicated the formation of the hypoiodite and the generation of the aldehyde but that the bicyclic alcohol had reformed to give the thermody-



Scheme 4. Synthesis of C-2 branched galactoside. Reagents used were the same as Scheme 2.



Scheme 5. Synthesis of 2-C-formyl glucosides. Reaction conditions were the same as Scheme 2.

namically stable bicyclic alcohol. The NMR of the photolyzed product showed H-2 as an apparent doublet of triplets with J = 5.7, 11.5 Hz. Thus H-2 should be nearly trans-diaxial to both H-3 and H-7, indicating 14 β as the epimerized product. Similar photochemically induced inversions of configuration of alcohols were reported by Binkley and co-workers [11].

The application of this formyl-transfer method to commercially available tri-O-acetyl-D-galactal was explored. The same two-step sequence of reactions (Scheme 2) was repeated to synthesize the radical precursor 23 (Scheme 4). Iodoetherification of the galactal gave only the diaxial galactoside 22 in 42% yield. Ozonolysis of 22 furnished the radical precursor 23 in 84% yield. Radical reaction of the galactoside 23 using tributyltin hydride conditions successfully produced the desired formyltransfer product 24 in 50% yield (0.05 M, 2 h) along with an inseparable mixture of what appeared to be the bicyclic alcohol 25 and the rearranged furanoside 26. The structures of these byproducts were not confirmed since the two compounds were inseparable.

The collective results of the radical reactions

indicated that the initial radical cyclization to the aldehvde occurred but the subsequent fragmentation of the alkoxyl radical to form the α -isopropoxy radical was problematic. Therefore, the intermediate α -isopropoxy radical was not stable enough to drive the reaction to completion. With this in mind, a modified radical substrate was synthesized to favor the fragmentation process by increasing the stability of the resulting radical. Initially, we tried changing the substrate such that the resulting radical from the fragmentation would be an α . α -diphenvl radical. In order to synthesize the radical substrate it was necessary to add α, α diphenyl allyl alcohol (easily prepared from benzophenone and vinylmagnesium bromide) to 5 and 6. However, many attempts at the iodoetherification reaction were unsuccessful presumably due to the low reactivity of the diphenyl allyl alcohol towards electrophilic reactions and also its propensity to rearrange to β-phenylcinnamyl alcohol with traces of acid. Next, α -phenyl allyl alcohol 27 was added to 5 and 6 since these substrates would produce a stable benzyl radical upon fragmentation (Scheme 5). The radical substrates 30 and 31 were prepared by reacting the glucals with α -phenyl allyl alcohol and N-iodosuccinimide in acetonitrile. The iodoetherification reaction furnished only the diaxial products as a 1:1 diastereomeric mixture at the allylic centers of the glucosides. Although the mixture of diastereomers were inseparable, the newly generated chiral center was not important since the radical reaction would produce a benzyl group. The mixture of diastereomers was ozonized to afford the key aldehydes 30 and 31 in 74% yields.

The modified radical precursor **30** yielded the formyl-transfer adduct **32** in 50% yield under optimal conditions. Radical reaction of the tri-*O*-benzyl iodoglucoside **31** produced the desired glucoside **33** in 84% yield with only traces of the glycal **18**. Therefore, the driving force of producing a benzyl radical caused the fragmentation to occur more efficiently, leading to higher yields of the desired C-2 formyl glucopyranosides. Thus, this simple three-step procedure of iodoetherification, ozonolysis, and radical cyclization–fragmentation efficiently converts glucals into α C-2 formyl pyranosides. To further explore this radical fragmentation process, this reaction was extended to nitrile-transfer reactions. Curran and co-workers reported several examples of intramolecular nitrile-transfer reactions on non-carbohydrate systems [12]. The radical substrates were prepared from **5** and **6** using acetone cyanohydrin and *N*-iodosuccinimide. This reaction successfully furnished **34** in 82% yield and **35** in 42% yield (Scheme 6). The iodoetherification reaction using mandelonitrile as the nucleophile gave unsatisfactory yields of the radical substrates, presumably due to the same reasons given for α, α -diphenyl allyl alcohol.

The iodoglucoside **34** successfully produced the nitrile-transfer product **36**, albeit in 30% yield along with the directly reduced glucoside **37**. The cyano-transfer reactions with the tri-*O*-benzyl glucoside **35** led to decomposition of the starting material.

A new method for the construction of C-2 branched sugars was developed using an intramolecular formyl- and cyano-transfer reactions via a radical cyclization-fragmentation process. The desired formyl-transfer product 11 was isolated in 41% yield along with the furanoside 16. Tris(trimethylsilyl)silane gave good yields of the formyl glycal 17, a compound otherwise difficult to synthesize directly. In addition, this study showed that the formyl-transfer method is a viable means to afford not only the α C-2 branched sugars but also the β C-2 branched sugar **21**. The yields of the formyl-transfer reactions were improved by changing the radical substrates to give a benzyl radical upon fragmentation. This method was also used to synthesize α C-2 branched galactopyranoside and α C-2 cyano glucoside 36.



Scheme 6. Synthesis of 2-C-cyanoglucoside.

3. Experimental

General methods.—¹H NMR spectra were recorded on a Bruker AF 200 spectrometer, operating at 200.132 MHz, a Bruker AM 360 spectrometer, operating at 360.134 MHz, and the Bruker ARX 400 spectrometer, operating at 400.130 MHz, and are so indicated. ¹³C NMR spectra were recorded on the AF 200. operating at 50.323 MHz, the AM 360, operating at 90.556 MHz, the ARX 400, operating at 100.625 MHz, and the ARX 500 spectrometer, operating at 125.767 MHz, and are also specified. IR spectra were recorded on a Nicolet 510 FTIR as a liquid film (neat) on sodium chloride plates, and the spectra were referenced to a polystyrene standard. High-resolution mass spectra were obtained from the Mass Spectrometry Facilities at the University of California, Riverside and at the University of California, Los Angeles using ZAB 7070 HP spectrometers. The chemical-impact highresolution mass spectra (CIHRMS) were obtained using ammonia as the reagent gas. All optical rotations were obtained from a Perkin-Elmer 243 polarimeter at ambient temperature and referenced to the sodium D line (589 nm). Thin-layer chromatography (TLC) was performed using E. Merck Silica Gel 60 F₂₅₄ 0.2 mm aluminum-backed plates. Visualization was accomplished using ultraviolet light or by using an anisaldehyde stain [p-anisaldehyde (2 mL) in EtOH (85 mL), glacial AcOH (10 mL), and concd H_2SO_4 (2 mL)]. Silica gel column chromatography was conducted on E. Merck Silica Gel 60 (70-230 mesh). Concentration in vacuo refers to the removal of solvent using a Büchi rotary evaporator.

1,1-Dimethyl-2-propenyl 3,4,6-tri-O-acetyl-2-deoxy-2-iodo- α -D-mannopyranoside (7). A solution of commercially available tri-O-acetyl-D-glucal (3 g, 11.0 mmol) in dry MeCN (30 mL) was stirred for 30 min with N-iodosuccinimide (2.5 g, 11.0 mmol) in the dark. To the reaction mixture was added dropwise 2methyl-3-buten-2-ol (5.7 mL, 55.0 mmol), and the mixture was stirred at rt for an additional 16 h. The reaction mixture was diluted with EtOAc, then washed several times with aq sodium thiosulfate until the organic layer

turned clear. The organic layer was combined, dried over MgSO₄ and concentrated in vacuo. The resulting yellow oil was purified by flash column chromatography (1:5 EtOAc-hexane) to give 3 g (84% based on recovered starting material) of the desired iodo sugar 7 as a clear liquid along with 1 g of recovered tri-O-acetyl-D-glucal: $[\alpha]_{D}$ + 18.3° (c 12.2, CH₂Cl₂); R_{f} 0.73 (1:1 EtOAc-hexane); IR (neat): 2982, 2940, 1752, 1433, 1416, 1368, 1231, 1113, 1030, 938, 920 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 5.81 (dd, 1 H, $J_{8,9}$ 10.7, $J_{8,9'}$ 17.5 Hz, H-8), 5.35 (t, 1 H, J_{3.5} 9.6 Hz, H-4), 5.28 (bs, 1 H, H-1), 5.19 (m, 2 H, H-9, H-9'), 4.68 (dd, 1 H, J_{2,3} 4.1, J_{3,4} 9.5 Hz, H-3), 4.40 (dd, 1 H, $J_{1,2}$ 1.3, $J_{2,3}$ 4.1 Hz, H-2), 4.23 (m, 3 H, H-5, H-6, H-6'), 2.09, 2.07, 2.05 (each s, 9 H, OAc), 1.35, 1.32 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 170.4, 169.6, 169.3, 142.0, 115.1, 96.4, 78.3, 68.9, 68.7, 67.6, 62.2, 31.4, 27.6, 24.9, 20.8, 20.6, 20.5; CIHRMS (m/z): 485.0672 (M⁺). Calcd for C₁₇H₂₅IO₈ 485.0641 (M⁺).

1,1-Dimethyl-2-propenyl 3,4,6-tri-O-benzyl-2-deoxy-2-iodo- α -D-mannopyranoside **(8**α). Prepared from 6 (1 g, 2.4 mmol) using dry THF in the manner as described for 7 to yield 460 mg (86% based on recovered starting material): $[\alpha]_{\rm D}$ + 12.2° (c 0.9, CH₂Cl₂); R_f 0.61 (1:2 EtOAc-hexane); IR (neat): 3031, 2979, 2926, 2861, 1497, 1455, 1364, 1144, 1113, 1053, 1028, 1000 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 7.30 (m, 15 H, Ph), 5.82 (dd, 1 H, $J_{8,9}$ 10.7, $J_{8,9'}$ 17.5 Hz, H-8), 5.36 (d, 1 H, $J_{1,2}$ 0.7 Hz, H-1), 5.16 (d, 1 H, H-9), 5.14 (d, 1 H, H-9'), 4.87 4.75, 4.72, 4.56, 5.11, 4.99 (each d, 6 H, J 10.5 Hz, benzyl), 4.37 (dd, 1 H, J_{1.2} 1.4, J_{2.3} 4.0 Hz, H-2), 4.07 (m, 1 H, H-5), 3.97 (dd, 1 H, J_{3,4} 8.8, J_{4,5} 9.7 Hz, H-4), 3.83 (dd, 1 H, $J_{5.6}$ 4.2, $J_{6.6'}$ 10.7 Hz, H-6), 3.69 (dd, 1 H, H-6'), 3.40 (dd, 1 H, J_{2.3} 4.0, J_{3.4} 8.7 Hz, H-3), 1.34, 1.28 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 142.7, 138.4, 138.2, 137.8, 128.3 (2C), 128.2, 128.1, 128.0, 127.7, 127.6 (2C), 127.3, 114.6, 96.7, 77.8, 76.9, 76.1, 75.2, 73.2, 71.9, 70.9, 68.8, 35.7, 27.8, 25.1; HRMS FAB (m/z): 627.1608 $(M^+ - H)$. Calcd for $C_{32}H_{36}IO_5$ 627.1581 (M⁺ – H).

1,1-Dimethyl-2-propenyl 3,4,6-tri-O-benzyl-2-deoxy-2-iodo- β -D-glucopyranoside (**8** β). Prepared from **6** (517 mg, 24 mmol) using MeCN in the manner described for 7 to yield 201 mg (62% based on recovered starting material): R_f 0.61 (1:2 EtOAc-hexane); ¹H NMR (360 MHz, CDCl₃): δ 7.29 (m, 15 H, Ph), 6.13 (dd, 1 H, $J_{8,9}$ 10.8, $J_{8,9'}$ 17.6 Hz, H-8), 5.22 (d, 1 H, H-9), 5.13 (d, 1 H, H-9'), 4.97 (d, 1 H, $J_{8,9}$ 10.1 Hz, benzyl), 4.85–4.78 (m, 2 H), 4.64–4.50 (m, 3 H), 4.59 (d, 1 H, $J_{1,2}$ 9.2 Hz, H-1), 3.92 (t, 1 H, $J_{1,3}$ 9.0 Hz, H-2), 3.74–3.58 (m, 3 H), 3.55 (t, 1 H, $J_{2,4}$ 9.7 Hz, H-3), 3.45 (m, 1 H), 1.41, 1.38 (each s, 6 H, Me).

1.1-Dimethyl-2-oxoethyl 3.4.6-tri-O-acetyl-2-deoxy-2-iodo- α -D-mannopyranoside (9). To a solution 7 (1.2 g, 2.6 mmol) in dry CH₂Cl₂ (120 mL) at -78 °C was added a steady stream of ozone until the clear solution turned dark blue (15 min). The excess ozone was purged with nitrogen (10 min) methyl sulfide (0.94 mL, 12.8 mmol) was added and the mixture was allowed to stir at -78 °C for an additional hour before warming to rt. The solvent was removed in vacuo and the resulting yellow residue was purified by flash column chromatography (3:7 EtOAc-hexane) to give 1.2 g (96%) of the desired aldehyde 9as a clear oil: $[\alpha]_{\rm D}$ + 22.2° (c 3.7, CH₂Cl₂); R_f 0.64 (1:1 EtOAc-hexane); IR (neat): 2986, 2940, 1748, 1435, 1370, 1231, 1161, 1115, 1040, 1013, 968, 949, 920 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 9.44 (s, 1 H, CHO), 5.28 (d, 1 H, J₁, 1.7 Hz, H-1), 5.25 (t, 1 H, J₃₅ 9.2 Hz, H-4), 4.60 (dd, 1 H, H-3), 4.47 (dd, 1 H, H-2), 4.14 (m, 2 H), 3.98 (m, 1 H), 2.07, 2.06, 2.03 (each s, 9 H, OAc), 1.34, 1.33 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 200.8, 170.5, 169.7, 169.3, 97.1, 82.1, 69.4, 68.7, 67.6, 62.2, 29.7, 21.5, 21.1, 20.8, 20.6, 20.5; CIHRMS (m/z): 504.0731 (M + NH₄⁺). Calcd for $C_{16}H_{27}INO_9$ 504.0751 (M + NH₄⁺).

1,1-Dimethyl-2-oxoethyl 3,4,6-tri-O-benzyl-2-deoxy-2-iodo-α-D-mannopyranoside (10α). Prepared from 8α (690 mg, 1.09 mmol) using the same procedure described for 9 to yield 0.60 g (86%): $[α]_D$ + 13.0° (*c* 1.5, CH₂Cl₂); *R_f* 0.69 (1:2 EtOAc-hexane); IR (neat): 2923, 2865, 1736, 1455, 1374, 1267, 1246, 1115, 1046, 911 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 9.54 (s, 1 H, CHO), 7.36 (m, 15 H, Ph), 5.46 (d, 1 H, *J*_{1,2} 1.58 Hz, H-1), 4.87, 4.75, 4.72 (each d, 3 H, *J* 10.1 Hz, benzyl), 4.56 (m, 4 H), 4.09 (m, 1 H), 3.97 (dd, 1 H, *J*_{3,4} 8.4, *J*_{4,5} 8.6 Hz, H-4), 3.80 (dd, 1 H, $J_{5,6}$ 4.6, $J_{6,6'}$ 10.7 Hz, H-6), 3.69 (dd, 1 H, H-6'), 3.41 (dd, 1 H, $J_{2,3}$ 4.0, $J_{3,4}$ 8.4 Hz, H-3), 1.34, 1.33 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 201.6, 138.2, 137.9, 137.5, 128.3 (2C), 128.2, 128.0 (2C), 127.8, 127.7, 127.5, 127.4, 97.3, 81.6, 76.7, 75.8, 75.1, 73.3, 72.5, 71.0, 68.7, 33.8, 21.6, 21.1; CIHRMS (m/z): 629.1415 (M⁺ – H). Calcd for C₃₁H₃₄IO₆ 629.1400 (M⁺ – H).

1,1-Dimethyl-2-oxoethyl 3,4,6-tri-O-benzyl-2-deoxy-2-iodo- β -D-glucopyranoside (10B).Prepared from 8β (52 mg, 0.083 mmol) using the same procedure described for 9 to yield 39.4 mg (76%): $[\alpha]_{D}$ + 28.2° (c 0.4, CH₂Cl₂); IR (neat): 3031, 2926, 2855, 1732, 1497, 1455, 1360, 1310, 1273, 1213, 1111, 1074, 1045, 1028, 911 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 9.66 (s, 1 H, CHO), 7.32 (m, 15 H, Ph), 4.97, 4.86, 4.79, 4.60, 4.58, 4.49 (each d, 6 H, J 10.1 Hz, benzyl), 4.68 (d, 1 H, $J_{1,2}$ 8.9 Hz, H-1), 3.95 (dd, 1 H, $J_{1,2}$ 8.9, J_{2.3} 9.1 Hz, H-2), 3.70 (m, 4 H), 3.42 (m, 1 H), 1.39, 1.34 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 202.5, 137.9, 137.7, 137.6, 128.5, 128.4 (2C), 128.1 (2C), 127.9, 127.8 (2C), 127.7, 98.3, 87.1, 82.5, 79.2, 75.3, 75.2 (2C), 73.5, 68.7, 32.6, 22.6, 19.2; (m/z): CIHRMS 648.1820 $(M + NH_4^+).$ Calcd for $C_{31}H_{34}INO_6$ 648.1822 (M + NH₄⁺).

Typical radical reaction using tributyltin hydride. To a refluxing solution of 9 (99.6 mg, 0.205 mmol) in degassed benzene (31 mL) was added a solution of tributyltin hydride (0.055 mL, 0.205 mmol) and AIBN (0.01 g, 0.061 mmol) over 2 h using a syringe pump. The reaction was allowed to heat to reflux for an additional 3 h then cooled to rt. The solvent was removed in vacuo, and the yellow residue was diluted with MeCN and washed with pentane several times to remove all the tin byproducts. The MeCN layers were combined and concentrated in vacuo to give a pale yellow oil. The products of the radical reaction were purified by flash column chromatography (3:7 EtOAc-hexane). Depending on the reaction conditions, i.e., the concentrations and the rate of addition of the tin hydride, we observed different products (see Tables 1 and 2).

Typical radical reaction using tris-(trimethylsilvl)silane (TTMSS). To a refluxing solution of 9 (236 mg, 0.484 mmol) in degassed toluene (15 mL) was added a solution of tris(trimethylsilyl)silane (0.15 mL. 0.484 mmol), and AIBN (0.04 g, 0.242 mmol) in toluene (5 mL) via syringe pump over a 1.5 h period. The reaction was allowed to heat at reflux for an additional 4 h before cooling to rt. The reaction mixture was concentrated in vacuo, then the products were purified by flash column chromatography (3:7 EtOAc-hexane). Depending on the reaction conditions, i.e., the concentrations and the rate of addition of the tin hydride, we observed different products (see Tables 1 and 2).

Isopropyl 3.4.6-tri-O-acetyl-2-deoxy-2-Cformyl- α -D-glucopyranoside (11): IR (neat): 2967, 1750, 1373, 1229, 1119, 1051 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.48 (d, 1 H, J_{2,CHO} 2.5 Hz, CHO), 5.81 (t, 1 H, J₃₅ 9.3 Hz, H-4), 5.25 (d, 1 H, $J_{1,2}$ 4.0 Hz, H-1), 5.03 (t, 1 H, J_{2.4} 9.7 Hz, H-3), 4.30 (dd, 1 H, J_{5,6} 4.5, J_{6,6'} 12.1 Hz, H-6), 4.15 (m, 1 H, H-5), 4.08 (dd, 1 H, H-6'), 3.88 (septet, 1 H, H-7), 2.78 (ddd, 1 H, H-2), 2.09, 2.05, 2.01 (each s, 9 H, OAc), 1.22, 1.11 (each d, 6 H, $J_{7.Me}$ 6.2 Hz, Me); ¹³C NMR (100 MHz, $CDCl_3$): δ 197.4, 170.6, 170.2, 169.8, 95.2, 77.1, 71.0, 68.9, 68.2, 68.1, 67.7, 61.9, 56.6, 23.0, 20.6 (2C); CIHRMS (m/z): 378.1761 $(M + NH_4^+)$. Calcd for $C_{16}H_{28}NO_9$ 378.1764 $(M + NH_4^+).$

Isopropyl 3,4,6-tri-O-benzyl-2-deoxy-2-Cformyl- α -D-glucopyranoside (12): R_{f} 0.73 (1:2) EtOAc-hexane); ¹H NMR (360 MHz, CDCl₃): δ 9.47 (d, 1 H, $J_{2 CHO}$ 2.6 Hz, CHO), 7.30 (m, 15 H, Ph), 5.19 (d, 1 H, J₁) 3.9 Hz, H-1), 4.90, 4.82, 4.77, 4.66, 4.56, 4.52 (each d, 6 H, J 11.0 Hz, benzyl), 4.36 (dd, 1 H, J_{3.4} 8.9, J_{4.5} 11.0 Hz, H-4), 3.94-3.65 (m, 5 H), 2.71 (ddd, 1 H, H-2), 1.16, 1.05 (each d, 6 H, $J_{7,Me}$ 6.2 Hz, Me); ¹³C NMR (100 MHz, CDCl₃): δ 199.8, 138.0, 137.8, 137.7, 128.4, 128.3, 128.2, 128.0, 127.8 (2C), 127.7, 127.6, 127.5, 95.4, 79.1, 74.9, 74.7, 73.5, 70.9, 69.8, 68.3, 58.5, 23.1, 21.2. (one aliphatic carbon not resolved); CIHRMS (m/z): 522.2832 $(M + NH_4^+).$ Calcd for $C_{31}H_{40}NO_6$ 522.2856 (M + NH₄⁺).

739

 $[4\mathbf{R} - (3\alpha\beta, 3a\beta, 4\beta, 5\alpha, 6\beta, 7a\beta)] - 6 - [(Acetyl$ oxy)methyl]hexahydro - 2,2,3 - trimethyl - 4Hfuro[2,3-b]pyran-4,5-diol, diacetate (13). Prepared from 7 (373 mg, 0.77 mmol) using typical radical reaction conditions described above to yield 113 mg (68% based on recovered starting material): R_f 0.44 (1:1 EtOAchexane); IR (neat): 2973, 1744, 1456, 1437, 1372, 1231, 1165, 1130, 1032, 963, 916 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 5.49–5.39 (m, 2 H), 5.30-5.29 (m, 1 H), 5.00-4.92 (m, 1 H), 4.92–5.00 (m, 1 H), 4.48–4.10 (m, 7 H), 3.15– 3.21 (m, 1 H, H-2), 2.52 (m, 1 H), 2.32 (m, 2 H), 2.10, 2.09, 2.07, 2.04, 2.03, 2.02 (each s, 18) H, OAc), 1.58, 1.52, 1.32, 1.30 (each s, 12 H, Me), 1.26, 0.94 (each d, 6 H, J_{7.Me} 6.8 Hz, Me); ¹³C NMR (100 MHz, CDCl₃): δ 170.7, 169.6 (2C), 97.3, 83.1, 70.9, 68.8, 67.3, 63.2, 48.0, 43.0, 27.5, 22.3, 20.9, 20.8, 20.7, 13.3; CIHRMS (m/z): 376.1976 $(M + NH_4^+)$. Calcd for $C_{17}H_{30}NO_8$ 376.1971 (M + NH₄⁺).

 $[3\mathbf{R} - (3\alpha, 3\alpha\beta, 4\beta, 5\alpha, 6\beta, 7\alpha\beta)] - 6 - [(Acetyloxy) - 6]$ methyl]hexahydro-2,2-dimethyl-4H-furo[2,3-b] pyran-3,4,5-triol, 4,5-diacetate (14). Data for 14 α : ¹H NMR (360 MHz, CDCl₃): δ 5.45 (d, 1 H, J_{1,2} 5.1 Hz, H-1), 5.39 (dd, 1 H, J_{3,4} 8.7, J_{4,5} 8.8 Hz, H-4), 5.15 (t, 1 H, J_{2.4} 8.9 Hz, H-3), 4.32–4.12 (m, 3 H), 2.34 (ddd, 1 H, H-2), 2.06, 2.05, 2.04 (each s, 9 H, OAc), 1.41, 1.21 (each s, 6 H, Me). Data for 14β : ¹H NMR (360 MHz, CDCl₃): δ 5.51 (d, 1 H, $J_{1,2}$ 5.5 Hz, H-1), 5.14 (t, 1 H, $J_{3,5}$ 6.4 Hz, H-4), 5.02 (dd, 1 H, J_{2,3} 6.5, J_{3,4} 9.2 Hz, H-3), 4.29 (dd, 1 H, J_{5.6} 5.1, J_{6.6'} 12.2 Hz, H-6), 4.13–3.99 (m, 3 H, H-5, H-6', H-7), 2.34 (ddd, 1 H, $J_{1,2}$ 5.7, $J_{2,7}$ 5.7, J_{2 3} 11.5 Hz, H-2), 2.08, 2.07, 2.06 (each s, 9 H, OAc), 1.44, 1.21 (each s, 6 H, Me). Data for the mixture of isomers $14\alpha\beta$: IR (neat): 3480, 2977, 2938, 1748, 1435, 1370, 1237, 1161, 1113, 1036, 916 cm⁻¹; ¹³C NMR (90 MHz, CDCl₃): δ 171.1, 170.8, 169.8, 169.5, 97.5, 97.4, 83.3, 82.1, 78.5, 76.6, 71.3, 69.8, 69.3, 68.5, 68.3, 68.2, 62.6, 62.4, 50.5, 48.2, 27.7, 23.0, 22.1, 20.9, 20.7 (2C); 27.8. CIHRMS (m/z): 378.1773 $(M + NH_4^+)$. Calcd for $C_{16}H_{28}NO_9$ 378.1764 (M + NH₄⁺).

4R-($3\alpha\beta$, $3a\beta$, 4β , 5α , 6β , $7a\beta$) Hexahydro-2,2dimethyl - 4,5 - bis(phenylmethoxy) - 6 - [(phenylmethoxy)methyl] - 4H - furo[2,3 - b]pyran - 3 - ol (15αβ). Data for 15α: R_f 0.50 (1:2 EtOAc– hexane); ¹H NMR (360 MHz, CDCl₃); δ 7.34

(m, 15 H, Ph), 5.19 (d, 1 H, $J_{1,2}$ 4.3 Hz, H-1), 4.63–4.32 (m, 7 H, benzyl, H-7), 4.04 (bs, 1 H, OH), 3.94 (dt, 1 H, J₂, 1.4, J₃₄ 7.4 Hz, H-3), 3.80 (t, 1 H, J 7.2 Hz), 3.60 (m, 3 H), 2.64 (ddd, 1 H, J_{1.2} 4.3, J_{2.7} 4.3, J_{2.3} 7.1 Hz, H-2), 1.36, 1.20 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 137.6, 136.5, 128.5 (2C), 128.4, 128.3, 128.1, 128.0, 127.8 (2C), 127.7, 127.6, 100.9, 87.9, 79.0, 76.5, 73.4, 72.3, 71.9, 71.1, 71.0, 68.3, 47.9, 28.9, 23.9. Data for **15**β: R_{c} 0.35 (1:2 EtOAc-hexane); IR (neat): 3445, 3031, 2930, 2867, 1497, 1455, 1366, 1275, 1207, 1111, 1067, 1028 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 7.31 (m, 15 H, Ph), 5.50 (d, 1 H, J₁, 4.9 Hz, H-1), 4.80, 4.78, 4.68, 4.56, 4.55, 4.43 (each d, 6 H, J 11.6 Hz, benzyl), 4.19 (dd, 1 H, J_{3,4} 7.7, J_{4,5} 7.7 Hz, H-4), 3.94 (dd, 1 H, J_{2,3} 8.2, J_{3,4} 8.2 Hz, H-3), 3.86 (m, 2 H), 3.68 (m, 2 H), 2.61 (ddd, 1 H, J₁₂ 5.7, J₂₇ 5.7, J_{2.3} 10.9 Hz, H-2), 2.60 (bs, 1 H, OH), 1.37, 1.19 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 138.2, 137.7, 128.5, 128.4, 128.3, 128.0, 127.9, 127.8 (2C), 127.7, 127.6, 98.4, 84.9, 77.2, 76.8, 75.6, 73.9, 73.4, 73.0, 69.4, 49.7, 27.8, 23.4, 1. (one aromatic and one aliphatic carbon not resolved); CIHRMS (m/z): 522.2856 $(M + NH_4^+).$ Calcd for $C_{31}H_{40}NO_6$ 522.2867 (M + NH₄⁺).

 $[3R - (3\alpha, 3a\beta, 4\beta, 5\beta(R^*), 6a\beta)] - 5 - [1, 2 - Bis -$ (acetyloxy)ethyl]hexahydro-2,2-dimethyl-furo-[2,3-b]furan-3,4-diol, 4-acetate (16). $[\alpha]_{D}$ -18.8° (c 0.8, CH₂Cl₂); R_f 0.36 (2:3 EtOAchexane); IR (neat): 3567, 2975, 2938, 1744, 1435, 1372, 1233, 1154, 1125, 1098, 1049, 984, 953 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 5.37 (bd, 1 H, H-3), 5.19 (d, 1 H, J_{1.2} 4.1 Hz, H-1), 4.74 (dd, 1 H, J_{3,4} 1.9, J_{4,5} 2.1 Hz, H-4), 4.08 (m, 2 H, H-6, H-6'), 4.05 (m, 1 H, H-5), 3.86 (dd, 1 H, J_{2.7} 6.1, J_{7.0H} 12.0 Hz, H-7), 2.46 (d, 1 H, OH), 2.45 (m, 1 H, H-2), 2.11, 2.10, 2.05 (each s, 9 H, OAc), 1.39, 1.19 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 170.6, 169.3, 169.1, 101.1, 87.7, 76.5, 70.2, 68.3, 66.5, 62.1, 47.2, 28.3, 23.5, 20.9, 20.8, 20.7; CIHRMS (m/z): 361.1488 $(M + H^+)$. Calcd for $C_{16}H_{25}O_9$ 361.1498 (M + H⁺).

1,5-Anhydro-3,4,6-tri-O-acetyl-2-deoxy-2-C-formyl-α-D-arabino-hex-1-enitol (17). $[α]_D$ 34.7° (c 0.5, CH₂Cl₂); R_f 0.30 (1:1 EtOAchexane); IR (neat): 2963, 2924, 2853, 1748, 1682, 1634, 1424, 1372, 1032, 1229, 1194, 1125, 1032 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 9.36 (bs, 1 H, CHO), 7.46 (s, 1 H, H-1), 5.68 (bd, 1 H, H-3), 5.18 (t, 1 H, $J_{3,5}$ 3.5 Hz, H-4), 4.62 (m, 1 H, H-5), 4.47 (dd, 1 H, $J_{5,6}$ 7.7, $J_{6,6'}$ 12.2 Hz, H-6), 4.17 (dd, 1 H, H-6'), 2.09, 2.06, 2.04 (each s, 9 H, OAc); ¹³C NMR (90 MHz, CDCl₃): δ 188.1, 170.3, 169.4, 169.2, 163.6, 115.7, 76.3, 65.6, 60.8, 60.6, 20.6; CIHRMS (m/z): 301.0931 (MH⁺). Calcd for C₁₃H₁₇O₈ 301.0923 (MH⁺).

1,5-Anhydro-3,4,6-tri-O-benzyl-2-deoxy-2-C-formyl- α -D-arabino-hex-1-enitol (18) [4a]. R_f 0.12 (1:2 EtOAc-hexane); ¹H NMR (360 MHz, CDCl₃): δ 9.41 (s, 1 H, CHO), 7.29 (m, 15 H, Ph), 4.75-4.41 (m, 9 H), 3.84-3.77 (m, 2 H), 3.62 (dd, 1 H, $J_{5,6}$ 4.6, $J_{6,6'}$ 10.8 Hz, H-6); CIHRMS (m/z): 445.2006 (MH⁺). Calcd for C₂₈H₂₉O₅ 445.2015 (MH⁺).

1,1-Dimethyl-2-oxoethyl 3,4,6-tri-O-acetyl-2-deoxy- α -D-mannopyranoside (19). IR (neat): 2978, 1746, 1437, 1370, 1231, 1152, 1123, 1090, 1047, 1005, 924 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 9.53 (s, 1 H, CHO), 5.37 (m, 1 H, H-3), 5.19 (bd, 1 H, J_{1.2} 3.3 Hz, H-1), 4.96 (t, 1 H, J_{3.5} 9.7 Hz, H-4), 4.27 (dd, 1 H, J_{5.6} 5.4, J_{6.6'} 12.0 Hz, H-6), 4.16 (m, 1 H, H-5), 3.98 (dd, 1 H, H-6'), 2.22 (dd, 1 H, $J_{1,2'}$ 5.3, $J_{22'}$ 12.8 Hz, H-2'), 2.08, 2.04, 2.01 (each s, 9 H, OAc), 1.87 (td, 1 H, $J_{1,2}$ 3.8, $J_{2,2'}$ 12.8 Hz, H-2), 1.33 (s, 6 H, 2Me); ¹³C NMR (90 MHz, $CDCl_3$): δ 201.8, 170.7, 170.3, 169.9, 92.6, 81.2, 69.5, 68.7, 68.2, 62.4, 35.6, 21.8, 20.9, 20.8, 20.7 (2C); CIHRMS (m/z): 378.1768 $(M + NH_4^+)$. Calcd for $C_{16}H_{28}NO_9$ 378.1764 $(M + NH_{4}^{+}).$

4R-(3αβ, 3aα, 4β, 5α, 6β, 7aβ) Hexahydro-2,2dimethyl - 4,5 - bis(phenylmethoxy) - 6 - [(phenylmethoxy)methyl] - 4H - furo[2,3 - b]pyran - 3 - ol (21). Data for one diastereomer: R_f 0.52 (1:2 EtOAc-hexane); IR (neat): 3463, 3032, 2924, 1497, 1455, 1366, 1321, 1076, 1028 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 7.30 (m, 15 H, Ph), 4.99 (d, 1 H, $J_{1,2}$ 3.4 Hz, H-1), 4.84, 4.78, 4.73, 4.69, 4.67, 4.54 (each d, 6 H, benzyl), 4.40 (d, 1 H, $J_{3,4}$ 9.8 Hz, H-3), 3.98 (m, 2 H), 3.81 (dd, 1 H, $J_{5,6}$ 3.6, $J_{6,6'}$ 11.0 Hz), 3.71 (dd, 1 H, H-6), 3.28 (m, 1 H), 2.71 (m, 1 H), 2.62 (bs, 1 H), 1.49, 1.19 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 138.4, 138.2, 137.7, 128.7, 128.5, 128.3, 128.1, 127.9, 127.8 (2C), 127.6, 127.5, 99.4, 80.7, 77.2, 76.1, 74.9, 74.1, 73.5 (2C), 68.8, 50.0, 29.5, 23.2. (one aliphatic carbon not resolved); CIHRMS (m/z): 522.2850 (M + NH₄⁺). Calcd for C₃₁H₄₀NO₆ 522.2856 (M + NH₄⁺).

1,1-Dimethyl-2-propenyl 3,4,6-tri-O-acetyl-2-deoxy-2-iodo- α -D-galactopyranoside (22).Prepared from commercially available tri-Oacetyl-D-galactal (1 mL, 4.52 mmol) in the same manner described for 7 to give 488 mg (42% based on recovered starting material): $[\alpha]_{\rm D}$ +46.1° (c 1.7, CH₂Cl₂); R_f 0.82 (2:3) EtOAc-hexane); IR (neat): 2978, 2928, 1750, 1497, 1455, 1370, 1231, 1105, 1028, 988, 930 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 5.81 (dd, 1 H, J_{7.8} 10.7, J_{7.8'} 17.5 Hz, H-7), 5.41 (bs, 1 H, H-1), 5.35 (m, 1 H), 5.20 (d, 1 H, H-8), 5.18 (d, 1 H, H-8'), 4.93 (dd, 1 H, J₂₃ 3.6, J₃₄ 4.2 Hz, H-3), 4.47 (ddd, 1 H, J 1.8, 6.5, 13.2 Hz), 4.12 (m, 3 H), 2.16, 2.05, 2.02 (each s, 9 H, OAc); ¹³C NMR (90 MHz, CDCl₃): δ 170.4, 170.1, 169.6, 142.3, 115.1, 98.1, 78.4, 66.4, 65.5, 65.3, 62.1, 27.8, 25.1, 23.4, 21.0, 20.9, 20.6; CIHRMS (m/z): 502.0938 (M + NH_{4}^{+}). Calcd for $C_{17}H_{29}INO_{8}$ 502.0957 (M + NH⁺).

1,1-Dimethyl-2-oxoethyl 3,4,6-tri-O-acetyl-2-deoxy-2-iodo- α -D-galactopyranoside (23).Prepared from 22 (218 mg, 0.45 mmol) using the same procedure described for 9 to give 184 mg (84%): $[\alpha]_{\rm D}$ + 61.3° (c 1.5, CH₂Cl₂); R_f 0.60 (2:3 EtOAc-hexane); IR (neat): 2982, 1752, 1464, 1435, 1373, 1231, 1169, 1109, 1040, 992, 955, 914 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 9.44 (s, 1 H, CHO), 5.38 (d, 1 H, J_{1.2} 1.9 Hz, H-1), 5.30 (dd, 1 H, J_{2.3} 2.8, $J_{3,4}$ 3.0 Hz, H-3), 4.92 (t, 1 H, $J_{3,5}$ 4.0 Hz, H-4), 4.39 (m, 1 H), 4.22 (m, 1 H), 4.11 (dd, 1 H, J_{5.6} 7.7, J_{6.6'} 11.6 Hz, H-6), 4.01 (dd, 1 H, H-6'), 2.07, 1.99, 1.96 (each s, 9 H, OAc), 1.26, 1.25 (each s, 6 H, Me); ¹³C NMR (50 MHz, CDCl₃): δ 201.1, 170.5, 170.0, 169.6, 98.2, 82.0, 67.5, 65.5, 61.9, 22.2, 21.8, 21.0, 20.9, 20.7. (Two aliphatic carbons not resolved); CIHRMS (m/z): 504.0731 (M + NH₄⁺). Calcd for $C_{16}H_{27}INO_9$ 504.0749 (M + NH₄⁺).

Isopropyl 3,4,6-tri-O-acetyl-2-deoxy-2-Cformyl- α -D-galactopyranoside (24). Prepared from 23 (59 mg, 0.12 mmol) using Bu₃SnH

radical reaction conditions (0.01 M, 3 h) to give 18.5 mg (43%) of **24** and 14 mg (32%) of a mixture of **25** and **26**: $[\alpha]_{\rm D}$ + 154.5° (c 0.62, CH₂Cl₂); IR (neat): 2977, 2930, 1750, 1437, 1372, 1233, 1163, 1125, 1071, 1017, 950, 928 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.48 (d, 1 H, J_{2,CHO} 2.7 Hz, CHO), 5.70 (dd, 1 H, J₃₄ 3.1, $J_{2,3}$ 11.8 Hz, H-3), 5.43 (bd, 1 H, H-4), 5.29 (d, 1 H, J_{1.2} 3.9 Hz, H-1), 4.31 (t, 1 H, J 6.9 Hz), 4.09 (m, 2 H), 3.87 (septet, 1 H, H-7), 2.95 (ddd, 1 H, H-2), 2.13, 2.04, 1.98 (each s, 9 H, OAc), 1.20, 0.92 (each d, 6 H, J_{7 Me} 6.1 Hz, Me); ¹³C NMR (90 MHz, CDCl₃): δ 198.9, 170.4, 170.2, 170.0, 95.8, 71.1, 66.9, 66.2, 66.0, 62.1, 51.4, 23.0, 21.4, 22.7, 20.6 (2C); CIHRMS (m/z): 378.1758 $(M + NH_4^+)$. Calcd for $C_{16}H_{28}NO_9$ 378.1764 (M + NH₄⁺).

 $4R - (3\alpha\beta, 3\alpha\beta, 4\beta, 5\alpha, 6\beta)] - 6 - [(Acetyloxy) - 6]$ methyl]hexahydro-2,2-dimethyl-4H-furo[2,3-b] pyran-3,4,5-triol, 4,5-diacetate (25), $4R-(3\alpha\beta)$, $3a\beta, 4\beta, 5\alpha(\mathbf{R}^*), 6a\beta)] - 5 - [1, 2 - bis(acetyloxy)$ ethyl]-hexahydro-2,2-dimethylfuro[2,3-b]furan-3,4-diol, 4-acetate (26). Data for the mixture of 25 and 26: IR (neat): 3470, 2926, 1748, 1462, 1433, 1372, 1231, 1047 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 5.66 (t, 1 H, J 3.7 Hz), 5.56 (d, 1 H, J 4.3 Hz), 5.35–3.31 (m, 3 H), 5.05 (dd, 1 H, J 3.0, 9.8 Hz), 4.65–3.92 (m), 2.74 (m, 1 H), 2.58 (m, 1 H), 2.11, 2.08, 2.07, 2.05, 2.04, 2.03 (each s, 18 H, OAc), 1.42, 1.27, 1.25, 1.23 (s, 12 H, Me); CIHRMS (m/ 378.1766 $(M + NH_4^+)$. Calcd for z): $C_{16}H_{28}NO_9$ 378.1764 (M + NH₄⁺).

1-Phenyl-2-propen-1-ol (27). To a stirred solution of benzaldehyde (1 g, 9.42 mmol) in dry tetrahydrofuran (50 mL) was added dropwise vinylmagnesium bromide (1.0 M in THF, 1.9 ml, 14.3 mmol) at 0 °C. The reaction mixture was allowed to warm to rt and stirred for an additional hour before quenching with aq NH₄Cl. The reaction mixture was concentrated, diluted with ether and washed with brine several times. The organic layer was combined, dried over MgSO₄ and concentrated in vacuo. Flash column chromatography (1:9 EtOAc-hexane) of the resulting oil gave 0.82 g (65%) of the desired alcohol 90 as a clear oil: R_f 0.52 (1:2 EtOAc-hexane); ¹H NMR (360 MHz, CDCl₃): δ 7.27 (m, 5 H, Ph), 5.99 (ddd, 1 H, J_{1,2} 6.0, J_{2,3} 10.3, J_{2,3'} 16.8 Hz, H-2), 5.27 (d, 1 H, H-3), 5.13 (d, 1 H, H-31), 5.12 (bd, 1 H, H-1), 2.23 (bs, 1 H, OH).

1-Phenyl-2-propenyl 3,4,6-tri-O-acetyl-2 $deoxy-2-iodo-\alpha$ -D-mannopyranoside (28). Prepared from 5 (28.8 g, 10.2 mmol) and 27 (1.5 g, 11.2 mmol) in the manner described for 7 to give 2.92 g (80% based on recovered starting material). Data for the mixture of diastereomers: R_f 0.36 (2:3 EtOAc-hexane); IR (neat): 3031, 2986, 2940, 1748, 1493, 1456, 1431, 1370, 1296, 1231, 1175, 1119, 1038, 945, 916 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 7.30 (m, 10 H, Ph), 5.97 (m, 1 H), 5.94 (m, 1 H), 5.42–5.16 (m, 11 H), 4.71 (dd, 1 H, J_{23} 4.2, J₃₄ 9.4 Hz, H-3), 4.61 (m, 1 H), 4.47 (dd, 1 H, $J_{1,2}$ 1.1, $J_{2,3}$ 4.4 Hz, H-2), 4.25–4.05 (m, 4 H), 3.89 (m, 12 H), 3.82 (m, 1 H), 2.12, 2.08, 2.06, 2.05, 2.02, 1.99 (each s, 18 H, OAc); ¹³C NMR (100 MHz, CDCl₃): δ 170.9, 170.5 (2C), 169.7, 169.4, 169.3, 139.6, 138.5, 137.7, 136.3, 128.7, 128.5, 128.4, 127.9, 127.2, 126.5, 118.6, 115.9, 99.0, 98.4, 80.3, 79.5, 69.3 (2C), 69.1, 69.0, 67.4, 67.2, 62.1, 61.8, 29.7, 29.6, 20.9, 20.8, 20.7, 20.6 (2C), 20.5; CIHRMS (m/z): 550.0920 (M + NH₄⁺). Calcd for $C_{21}H_{29}INO_8$ 550.0938 (M + NH₄⁺).

1-Phenyl-2-propenyl 3,4,6-tri-O-benzyl-2 $deoxy-2-iodo-\alpha$ -D-mannopyranoside (29). Prepared from 6 (2.1 g, 4.9 mmol) and 27 (0.73 g, 5.42 mmol) in the manner described for 7 to yield 2 g (60%). Data for the mixture of diastereomers: R_f 0.68 (1:6 EtOAc-hexane); IR (neat): 3088, 3063, 3031, 2865, 1605, 1497, 1455, 1362, 1308, 1291, 1265, 1208, 1113, 1028, 932, 912 cm⁻¹; ¹H NMR (360 MHz, $CDCl_3$): δ 7.36 (m, 40 H, Ph), 5.98 (dd, 1 H, J 6.1, 10.4 Hz), 5.95 (dd, 1 H, J 6.2, 10.3 Hz), 5.84 (dd, 1 H, J 7.7, 10.1 Hz), 5.79 (dd, 1 H, J 7.7, 9.9 Hz), 5.48–4.47 (m, 19 H), 3.98 (m, 4 H), 3.87–3.58 (m, 5 H), 3.46 (dd, 1 H, J 4.1, 8.2 Hz), 3.36 (dd, 1 H, J 4.1, 8.7 Hz); ¹³C NMR (90 MHz, CDCl₃): δ 140.2, 139.3, 138.5, 138.3, 138.2, 137.9, 137.8, 136.7, 128.5 (2C), 128.4 (2C), 128.3 (2C), 128.2 (2C), 127.9 (2C), 127.8, 127.7, 127.6, 127.5, 126.8, 126.7, 118.8, 115.8, 99.3, 99.0, 98.8, 85.9, 80.4, 79.5, 78.9, 77.6, 77.2, 76.2, 75.5 (2C), 75.4, 75.3, 74.9, 73.5, 73.4 (2C), 72.6 (2C), 71.1 (2C), 68.9, 68.8, 68.6, 60.4, 34.0, 33.9, 21.1, 14.3. (Several carbons not resolved); CIHRMS (m/z): 675.1627 (M – H). Calcd for $C_{36}H_{36}IO_5$ 675.1608 (M – H).

1-Phenyl-2-oxoethyl 3,4,6-tri-O-acetyl-2 $deoxy-2-iodo-\alpha$ -D-mannopyranoside (30). Prepared from 28 (0.10 g, 0.19 mmol) using the same procedure described for 9 to yield 76 mg (74%): IR of mixture (neat): 2937, 1746, 1494, 1455, 1370, 1232, 1124, 1061, 913 cm⁻¹; Data for one of the diastereomers: ¹H NMR (360 MHz, CDCl₃): δ 9.61 (bs, 1 H, CHO), 7.42 (m, 5 H, Ph), 5.46 (bs, 1 H, H-1), 5.38 (t, 1 H, J₃₅ 9.8 Hz, H-4), 5.11 (bs, 1 H, H-7), 4.81 (dd, 1 H, J_{1.2} 1.2, J_{2.3} 4.4 Hz, H-2), 4.65 (dd, 1 H, J_{2.3} 4.4, J_{3.4} 9.6 Hz, H-3), 4.06 (dd, 1 H, H-6), 3.89 (dd, 1 H, H-6'), 3.77 (ddd, 1 H, H-5), 2.09, 2.07, 2.00 (each s, 9 H, OAc); ¹³C NMR (90 MHz, CDCl₃): δ 196.3, 170.5, 169.8. 169.4, 132.7, 129.4, 129.3, 127.7, 101.3, 83.6, 70.0, 68.7, 67.3, 61.8, 28.5, 20.9, 20.7, 20.6. Data for one of the diastereomers: ¹H NMR (360 MHz, CDCl₃): δ 9.64 (d, 1 H, J 1.7 Hz, CHO), 7.42 (m, 5 H, Ph), 5.40 (t, 1 H, J_{3.5} 9.4 Hz, H-4), 5.20 (d, 1 H, J 1.4 Hz, H-1 or H-7), 4.99 (d, 1 H, J 1.4 Hz, H-1 or H-7), 4.77 (dd, 1 H, J_{2.3} 4.4, J_{3.4} 9.3 Hz, H-3), 4.62 (dd, 1 H, J_{1.2} 1.5, J_{2.3} 4.3 Hz, H-2), 4.21 (m, 3 H, H-5, H-6, H-6'), 2.13, 2.10, 2.09 (each s, 9 H, OAc); ¹³C NMR (90 MHz, CDCl₃): δ 195.8, 170.6, 169.8, 169.5, 132.3, 129.6, 129.4, 127.7, 99.6, 83.8, 70.0, 69.0, 67.3, 62.1, 28.4, 20.9, 20.7, 20.6; CIHRMS (m/z): 552.0720 $(M + NH_4^+)$. Calcd for $C_{20}H_{27}INO_9$ 552.0731 (M + NH₄⁺).

1-Phenyl-2-oxoethyl 3,4,6-tri-O-benzvl-2 $deoxy-2-iodo-\alpha$ -D-mannopyranoside (31). Prepared from 29 (570 mg, 0.84 mmol) using the same procedure described for 9 to yield 425 mg (74%). Data for one of the diastereomers: ¹H NMR (360 MHz, CDCl₃): δ 9.60 (s, 1 H, CHO), 7.36 (m, 20 H, Ph), 5.54 (bs, 1 H), 5.21 (s, 1 H), 4.85, 4.78, 4.68, 4.58, 4.50, 4.49 (each d, 6 H, J 10.6 Hz, benzyl), 4.82 (dd, 1 H, J 1.2, 4.2 Hz), 3.95 (t, 1 H, J 9.4 Hz), 3.73 (m, 2 H), 3.58 (m, 1 H), 3.37 (dd, 1 H, J 4.1, 8.7 Hz). Data for one of the diastereomers: ¹H NMR (360 MHz, CDCl₃): δ 9.60 (d, 1 H, J 1.4 Hz, CHO), 7.36 (m, 20 H, Ph), 5.26 (d, 1 H, J 1.1 Hz), 5.04 (d, 1 H, J 1.5 Hz), 4.89, 4.76, 4.74, 4.61, 4.56, 4.52 (each d, 6 H, J 10.7 Hz, benzyl), 4.58 (d, 1 H, J 1.6 Hz), 4.02 (m, 2 H), 3.82 (dd, 1 H, J_{5.6} 4.5, J_{6.6'} 10.9 Hz, H-6), 3.75 (dd, 1 H, J 1.5, 10.9 Hz), 3.51 (dd, 1 H, J 4.1, 8.2 Hz). Data of the mixture of diastereomers: IR (neat): 3031, 2924, 1734,

1497, 1455, 1362, 1267, 1101, 1028 cm⁻¹; ¹³C NMR (90 MHz, CDCl₃): δ 197.0, 196.4, 138.2, 138.0, 137.9, 137.5, 132.6, 129.3, 129.2, 129.1 (2C), 128.4 (3C), 128.3(2C), 128.2, 128.1 (2C), 128.0, 127.8 (2C), 127.6 (2C), 127.5 (2C), 101.1, 99.6, 83.2, 82.7, 76.8, 76.5, 75.8, 75.7, 75.4, 75.3, 73.4, 73.3, 73.0, 72.8, 71.2, 71.0, 68.6, 68.5, 32.5, 29.7. (Several carbons not resolved); CIHRMS (*m*/*z*): 696.1805 (M + NH₄⁺). Calcd for C₃₅H₃₉INO₆ 696.1822 (M + NH₄⁺).

Benzyl 3,4,6-tri-O-acetyl-2-deoxy-2-C-for $myl-\alpha$ -D-glucopyranoside (32). Prepared from 30 (75 mg, 0.14 mmol) using Bu₃SnH (0.046 mL, 0.17 mmol) radical reaction conditions (0.60 M, 3 h) to yield 28.5 mg (50%): $[\alpha]_{D}$ $+ 141.9^{\circ}$ (c 0.0016, CH₂Cl₂); IR (neat): 3034, 2957, 1748, 1455, 1433, 1370, 1229, 1125, 1024, 922 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.51 (d, 1 H, $J_{2,CHO}$ 2.4 Hz, CHO), 7.33 (m, 5 H, Ph), 5.81 (t, 1 H, J_{3.5} 9.7 Hz, H-4), 5.23 (d, 1 H, $J_{1,2}$ 3.9 Hz, H-1), 5.05 (t, 1 H, $J_{2,4}$ 9.7 Hz, H-3), 4.68, 4.52 (each d, 2 H, J 11.9 Hz, benzyl), 4.30 (dd, 1 H, J_{5.6} 4.2, J_{6.6'} 12.1 Hz, H-6), 4.06 (m, 2 H, H-5, H-6'), 2.84 (ddd, 1 H, J_{2,CHO} 2.7, J_{1,2} 3.6, J_{2.3} 8.4 Hz, H-2), 2.11, 2.04, 2.00 (each s, 9 H, OAc); ¹³C NMR (125 MHz, CDCl₃): δ 196.8, 170.5, 170.1, 169.7, 135.9, 128.5, 128.2, 128.0, 95.7, 69.8, 68.7, 67.9, 61.8, 56.6, 30.8, 20.6 (2C), 20.5; CIHRMS (m/z): 426.1752 $(M + NH_4^+)$. Calcd for $C_{20}H_{28}NO_9$ 426.1764 (M + NH₄⁺).

Benzyl 3,4,6-tri-O-benzyl-2-deoxy-2-C-for $myl-\alpha$ -D-glucopyranoside (33). Prepared from **31** (76.5 mg, 0.112 mmol) using Bu₂SnH (0.03 mL, 0.112 mmol) radical reaction conditions (0.01 M, 3 h) to yield 26 mg (84% based on recovered starting material): $[\alpha]_D + 96.6^\circ$ (c 0.4, CH₂Cl₂); R_f 0.52 (1:2 EtOAc-hexane); IR (neat): 3032, 2924, 2853, 1727, 1497, 1455, 1360, 1273, 1125, 1059, 1026 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 9.51 (d, 1 H, $J_{2,CHO}$ 2.6 Hz, CHO), 7.35–7.17 (m, 20 H, Ph), 5.20 (d, 1 H, J_{1,2} 3.9 Hz, H-1), 4.90–4.45 (m, 8 H, benzyl), 4.39 (dd, 1 H, J_{3,4} 8.8, J_{4,5} 8.9 Hz, H-4), 3.90–3.87 (m, 1 H), 3.78 (m, 2 H), 3.66 (dd, 1 H, J 1.9, 10.6 Hz), 2.78 (ddd, 1 H, $J_{2 \text{ CHO}}$ 2.6, $J_{1,2}$ 3.8, $J_{2,3}$ 6.5 Hz, H-2); ¹³C NMR (90 MHz, $CDCl_3$): δ 199.4, 138.2, 138.0, 137.8, 136.9, 128.5 (2C), 128.4 (2C), 127.9, 127.8 (3C), 96.5, 79.1, 76.5, 75.0, 74.9, 73.6, 71.2, 69.3, 68.3, 58.6. (Several aromatic carbons not resolved); CIHRMS (m/z): 551.2448 (M⁺ – H). Calcd for C₃₅H₃₅O₆ 551.2434 (M⁺ – H).

1-Cvano-1-methylethyl 3,4,6-tri-O-acetyl-2 $deoxy-2-iodo-\alpha$ -D-mannopyranoside (34). Prepared from 5 (2 g, 7.35 mmol) and acetone cyanohydrin (2.68 mL, 29.4 mmol) in the manner described for 7 to yield 1.4 g (82%) based on recovered starting material): $[\alpha]_{D}$ $+4.26^{\circ}$ (c 10.8, CH₂Cl₂); R_c 0.48 (1:1 EtOAchexane); IR (neat): 2994, 2963, 2942, 2200, 1748, 1435, 1372, 1221, 1167, 1119, 1084, 1032, 1007, 916 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 5.55 (d, 1 H, $J_{1,2}$ 1.2 Hz, H-1), 5.37 (t, 1 H, $J_{3,5}$ 9.6 Hz, H-4), 4.56 (dd, 1 H, $J_{2,3}$ 4.3, $J_{3.4}$ 9.3 Hz, H-3), 4.46 (dd, 1 H, $J_{1,2}$ 1.5, J_{23} 4.3 Hz, H-2), 4.23–4.10 (m, 3 H, H-5, H-6, H-6'), 2.07, 2.06, 2.03 (each s, 9 H, OAc), 1.64, 1.63 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 170.5, 169.8, 169.3, 120.0, 98.5, 70.5, 70.1, 68.6, 67.2, 61.8, 28.8, 27.9, 26.6, 20.9, 20.6 (2C); CIHRMS (m/z): 501.0716 (M + NH₄⁺). Calcd for $C_{16}H_{26}IN_2O_8$ $501.0734 (M + NH_4^+).$

1-Cvano-1-methylethyl 3,4,6-tri-O-benzyl-2-deoxy-2-iodo- α -D-mannopyranoside (35).Prepared from 6 (1.22 g, 2.93 mmol) and acetone cyanohydrin (1.5 mL, 14.7 mmol) in the manner described for 7 to yield 0.78 g (42%): $[\alpha]_{\rm D}$ + 20.0° (c 1.0, CH₂Cl₂); R_f 0.43 (1:2 EtOAc-hexane); IR (neat): 3088, 3063, 3031, 2992, 2919, 2867, 1497, 1455, 1364, 1293, 1267, 1208, 1171, 1113, 1028, 999 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 7.43–7.15 (m, 15 H, Ph), 5.65 (bs, 1 H, H-1), 4.85 (d, 1 H, J 10.5 Hz, benzyl), 4.76–4.70 (m, 2 H), 4.55– 4.44 (m, 4 H), 4.04–3.95 (m, 1 H), 3.82 (dd, 1 H, J_{2.3} 3.4, J_{3.4} 10.8 Hz, H-3), 3.73 (dd, 1 H, J 1.5, 11.0 Hz), 3.28 (dd, 1 H, J 4.1, 7.6 Hz), 1.63, 1.60 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 138.2, 138.0, 137.5, 128.4 (2C), 128.3, 128.1, 128.0, 127.9, 127.8, 127.6, 127.5, 120.4, 98.9, 76.5, 75.6, 75.3, 73.4, 73.2, 71.0, 70.4, 68.3, 32.8, 27.7, 27.2; CIHRMS (m/z): 645.1841 $(M + NH_4^+).$ Calcd for $C_{31}H_{38}IN_2O_5$ 645.1826 (M + NH₄⁺).

1-Methylethyl 3,4,6-tri-O-acetyl-C-cyano-2deoxy-2- α -D-glucopyranoside (**36**). Prepared from **34** (122 mg, 0.25 mmol) using Bu₃SnH (0.08 mL, 0.28 mmol) radical reaction conditions (0.13 M, 2 h) to yield 26 mg (30%) of **36**

and 26 mg (30%) of **37**: $[\alpha]_{\rm D}$ + 187.5° (*c* 0.08, CH₂Cl₂); IR (neat): 2977, 2932, 2255, 1752. 1456, 1372, 1227, 1119, 1047, 959, 924 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 5.58 (t, 1 H, J_{2,4} 9.3 Hz, H-3), 5.20 (d, 1 H, J_{1,2} 3.3 Hz, H-1), 4.93 (t, 1 H, J_{3,5} 9.8 Hz, H-4), 4.27 (dd, 1 H, $J_{5,6}$ 4.5, $J_{6,6'}$ 12.3 Hz, H-6), 4.13 (ddd, 1 H, J 2.2, 4.4, 10.2 Hz), 4.06 (m, 1 H), 3.98 (septet, 1 H, H-7), 3.07 (dd, 1 H, $J_{1,2}$ 3.4, $J_{2,3}$ 11.3 Hz, H-2), 2.04, 2.03, 2.01 (each s, 9 H, OAc), 1.27, 1.25 (each d, 6 H, J_{7 Me} 6.1 Hz, Me); ¹³C NMR (100 MHz, CDCl₃): δ 170.4, 169.6, 169.4, 115.1, 94.0, 77.1, 72.3, 68.3 (2C), 67.8, 61.6, 38.8, 30.8, 22.9, 21.3, 20.5; CIHRMS (m/z): 375.1763 $(M + NH_4^+)$. Calcd for $C_{16}H_{27}N_2O_8$ 375.1767 (M + NH₄⁺).

1-Cyano-1-methylethyl 3,4,6-tri-O-acetyl-2deoxy-α-D-glucopyranoside (**37**). IR (neat): 2994, 2963, 1746, 1441, 1370, 1304, 1233, 1173, 1154, 1123, 1088, 1049, 1017, 1003 cm⁻¹; ¹H NMR (360 MHz, CDCl₃): δ 5.42 (bd, 1 H, $J_{1,2}$ 2.2 Hz, H-1), 5.30 (m, 1 H, H-3), 5.02 (t, 1 H, $J_{3,5}$ 9.8 Hz, H-4), 4.34 (dd, 1 H, $J_{5,6}$ 4.2, $J_{6,6'}$ 12.2 Hz, H-6), 4.13–4.04 (m, 2 H, H-5, H-6'), 2.19 (m, 1 H, H-2), 2.07, 2.04, 2.01 (each s, 9 H, OAc), 1.91 (m, 1 H, H-2'), 1.65, 1.64 (each s, 6 H, Me); ¹³C NMR (90 MHz, CDCl₃): δ 170.6, 170.2, 169.8, 120.5, 94.2, 70.0, 69.0, 68.9, 68.5, 61.9, 35.4, 28.1, 26.7, 20.9, 20.7 (2C); CIHRMS (m/z): 375.1758 (M + NH₄⁺). Calcd for C₁₆H₂₇N₂O₈ 375.1767 (M + NH₄⁺).

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