- Chow, R. Serlin, and C. E. Strouse, J. Am. Chem. Soc., 97, 7230 (1975). (6) (a) S. G. Boxer and G. L. Closs, *J. Am. Chem. Soc.*, **98**, 5405 (1976); (b)
- S. G. Boxer, Ph.D. Dissertation, Department of Chemistry, University of Chicago, Chicago, III., 1976.
- .. Shipman, T. M. Cotton, J. R. Norris, and J. J. Katz, Proc. Natl. Acad. Sci. U.S.A., 73, 1791 (1976); (b) M. R. Wasielewski, M. H. Studier, and J. J. Katz, ibid., 73, 4282 (1976).
- T. M. Cotton, Ph.D. Dissertation, Department of Chemistry, Northwestern University, 1976.
- (9) R. H. Clarke and D. R. Hobart, FEBS Lett., 82, 155 (1977).
- (10) J. Breton, *Biochim. Biophys. Acta*, **453**, 66 (1977).
 (11) (a) F. K. Fong and N. Winograd, *J. Am. Chem. Soc.*, **98**, 2287 (1976); (b) L. M. Fetterman, L. Galloway, N. Winograd, and F. K. Fong, ibid., 99, 653
- (12) The electron spin resonance spectroscopic observation of the photooxidation of (ChI a·H₂O)₂ and (ChI a·2H₂O)₂ by water was reported in ref 4j, in which the biological significance of (ChI a·H₂O)₂ was also summarized. The proposal that (ChI a·2H₂O)₂ is the water splitting reaction center aggregate was suggested by this ESR observation⁴) and by the pH dependence of the photogalvanic response arising from (ChI a·2H₂O)_n—water light reactions.41 The in vitro photochemical splitting of water by (Chl a-2H2O)n was established by the observation of gaseous evolution of H₂, HD, D₂, ¹⁸O18O, and ¹⁸O₂ by mass spectroscopy and by pyrolysis. ^{4k,l} in the in vitro approach to photosynthesis research, ^{4l} the established properties of in vivo reaction center ChI a aggregates are compared with those of in vitro molecular adducts whose intermolecular interactions and photochemical behavior can be characterized under controlled laboratory conditions.
- (13) For a review of recent developments, see F. K. Fong, Photochem. Photobiol., submitted.
- (14) R. Willstätter and A. Stoll, "Investigations on Chlorophyll", F. M. Schertz and A. R. Merg, Translators, the Science Press Printing Co., Lancaster, Pa., 1928, p 31.
- (15) R. Livingston, in "Photosynthesis in Plants", Iowa State College Press, Ames, Iowa, 1949.
- (16) R. Livingston, W. F. Watson, and J. McArdle, J. Am. Chem. Soc., 71, 1542
- (1949). (17) È. Rabinowitch, in "Photosynthesis I", Interscience, New York, N.Y., 1945,

- pp 450–451.
 (18) A. S. Holt and E. E. E. Jacobs, *Am. J. Bot.*, **41**, 710 (1954).
 (19) K. Ballschmiter and J. J. Katz, *J. Am. Chem. Soc.*, **91**, 2661 (1969).
 (20) F. A. Cotton and G. Wilkinson, "Advanced Inorganic Chemistry", 3rd ed, Wiley-Interscience, New York, N.Y., 1972, pp 218, 219.

- (21) J. J. Katz and J. R. Norris, Curr. Top. Bioenerg., 5, 41 (1973).
- (22) See, for example, L. L. Shipman and J. J. Katz, J. Phys. Chem., 81, 577 (1977)
- H. H. Strain and W. A. Svec, in "Chlorophylls", L. P. Vernon and G. R. Seely, Ed., Academic Press, New York, N.Y., 1966, p 21.
- (24) (a) J. R. Wilson and M. D. Nutting, Anal. Chem., 35, 144 (1963); (b) L. P. Vernon, ibid., 32, 1144 (1960).
- (25) K. S. Kim, W. E. Baitinger, J. W. Amy, and N. Winograd, J. Electron Spectrosc. Relat. Phenom., 5, 351 (1974).
- D. Karweik, N. Winograd, D. G. Davis, and K. M. Kadish, J. Am. Chem. Soc., 96. 591 (1974)
- D. Karweik and N. Winograd, *Inorg. Chem.*, **15**, 2336 (1976). Y. Niwa, H. Kobayashi, and T. Tsuchiya, *J. Chem. Phys.*, **60**, 799 (1974)
- (29) S. G. Boxer, G. L. Closs, and J. J. Katz, J. Am. Chem. Soc., 96, 7058
- (30) O. Warburg and G. Krippahl, Z. Naturforsch. B, 11, 179 (1956).
- J. Franck, in "Research on Photosynthesis", H. Gaffron, Ed., Interscience, New York, N.Y., 1957, p 142.
 (32) D. Mauzerall, *Philos. Trans. R. Soc. London, Ser. B*, **273**, 287 (1976).
- A. A. Krasnovsky, K. K. Voinovskaya, and L. M. Kosobutskaya, Dokl. Akad. Nauk SSSR, 85, 389 (1952).
- A. A. Kransovsky and L. M. Kosobutskaya, Dokl. Akad. Nauk SSSR, 91, 343 (1953).
- (35) S. S. Brody, Science, 128, 838 (1958).
- S. S. Brody, and M. Brody, *Biochim. Biophys. Acta*, **54**, 495 (1961). W. D. Bellamy, G. L. Gaines, Jr., and A. G. Tweet, *J. Chem. Phys.*, **39**, 2528 (1963)
- (38) E. E. Jacobs and A. S. Holt, J. Chem. Phys., 22, 142 (1954).
- B. Ke, in ref 23, pp 253-279.
- G. Sherman and S. F. Wang, Nature (London), 211, 588 (1966).
- (41) D. Mauzerall, Encycl. Plant Physiol., New Ser., 5, 117–124 (1977)
- (42) D. Mauzerall and G. Feher, Biochim. Biophys. Acta, 79, 430 (1964)
- (43) F. Hong and D. Mauzerall, Proc. Natl. Acad. Sci. U.S.A., 71, 1564
- P. Carapellucci and D. Mauzerall, Ann. N.Y. Acad. Sci., 244, 214 (1975).
- (45) J. H. Fuhrhop, Z. Naturforsch. B, 25, 255 (1970).
- J. H. Fuhrhop and D. Mauzerall, J. Am. Chem. Soc., 91, 4174 (1969). L. Galloway, J. Roettger, D. R. Fruge, and F. K. Fong, J. Am. Chem. Soc., 100, 4635 (1978).
- R. P. H. Kooyman, T. J. Schaafsma, and J. F. Kleibeuker, Photochem. Photobiol., 26, 235 (1977).

Communications to the Editor

Total Synthesis of (\pm) -Seychellene. Use of Silyloxydienes in Synthesis¹

Seychellene (1), a plant sesquiterpene found in commercial patchouli oil and used in perfumery,² has been a common synthetic target due to its novel tricyclic structure. Several fine syntheses of seychellene have been published³⁻⁶ which range in yield from extremely low to \simeq 7%. We wish to report now a very direct and efficient total synthesis of seychellene (1) in only 10 steps, which demonstrates the very high synthetic utility of 2-silyloxydienes^{7,8} in Diels-Alder approaches to polycyclic natural products.

In our approach we chose to build the necessary bicyclo[2.2.2] octane system in the first step of the synthesis with appropriate functionality so positioned that the final ring closure could be simply accomplished. To this end the 2-trimethylsilyloxycyclohexadiene (3), which was prepared in 95% yield (LDA, THF, -78 °C/Me₃SiCl) from the readily available 2,3-dimethylcyclohexenone (2), was reacted with methyl vinyl ketone 4 (neat, sealed tube, 110 °C, 18 h) to afford a 79% yield of the Diels-Alder adduct 5 (oil; bp 80 °C (0.2 mm); NMR (CCl₄) δ 1.90 (s, 3 H), 1.50 (s, 3 H), 1.07 (s, 3 H), 0.13 (s, 9 H); IR (liquid film) 1700, 1670 cm⁻¹) (Scheme I). We assigned the structure 5, namely with the acetyl group endo and 1,4 to the trimethylsilyloxy function, to this Diels-Alder product on the basis of analogy to our earlier work⁷ and to that of others⁸ which had shown that the trimethylsilyloxy group was a very good director in such cycloaddition reactions. Indirect evidence for this assignment was obtained when the silyl enol ether of 5 was hydrolyzed in acid whereupon only dione 6a (NMR (CCl₄) δ 2.12 (s, 3 H), 1.11 (s, 3 H), 1.00 (d, 3 H, J = 7 Hz); IR (liquid film) 1710 cm⁻¹) was formed, and in base where a mixture of 6a and a more stable isomer 6b (NMR

Scheme I

(CCl₄) δ 2.12 (s, 3 H), 0.97 (d, 3 H, J = 6 Hz), 0.95 (s, 3 H); IR (liquid film) 1710 cm⁻¹) was produced. Pure dione **6a** could also be converted to a mixture in which **6b** predominated by base treatment. We assumed that **6a** and **6b** are the endo (syn to carbonyl) and exo (anti to carbonyl) isomers, respectively. However, the correctness of this assignment was not proven until the completion of the synthesis when the synthetic product could be compared with authentic material.

The next step of the synthesis involved the addition of the final two carbon atoms to the side chain. This would effectively complete the sesquiterpene skeleton since the introduction of the methylene carbon of 1 from the ketone was well known.³ The use of the silvloxydiene in the initial Diels-Alder greatly facilitated this next step since it provided directly a dione monoprotected at the desired carbonyl. All attempts at twocarbon functionalization of the ketone by means of Wittig, Wadsworth-Emmons, or Reformatskii reactions failed, possibly owing to the crowded steric environment of the carbonyl of 5. However, we were successful in adding vinylmagnesium bromide to 5 to furnish, after aqueous acid workup, the hydroxy ketone 7 (NMR (CCl₄) δ 4.9-6.4 (m, 3 H, vinyl pattern), 1.27 (s, 3 H), 1.03 (s, 3 H), 0.96 (d, 3 H, J = 7 Hz); IR (liquid film) 3400, 1700 cm, 1660 cm⁻¹) as a mixture of diastereomers in 73% yield.10

Transposition of the allylic alcohol function was easily accomplished by treatment of **7** with acetic acid containing a drop of sulfuric acid to give the acetate **8**, which could be hydrolyzed in base to the alcohol **9** (NMR (CCl₄) δ 5.38 (t, 1 H, J = 7 Hz), 4.07 (d, 2 H, J = 7 Hz), 0.97 (d, 3H, J = 7 Hz), 0.80 (s, 3 H); IR (liquid film) 3300, 1710, 1650 cm⁻¹), thus available from **7** in quantitative yield (Scheme II). ¹⁰ By analogy to the work of Schmalzl, ⁴ who reported that reduction of the acrylate **10** gave a mixture of alcohols **11a** and **11b** in which the desired

isomer 11a greatly predominated, we expected that reduction of 9 should afford the desired isomer 12a as the major isomer. In reality, a 1:1 mixture of the two alcohols 12a and 12b was produced upon catalytic hydrogenation over a rhodium-onalumina catalyst. The use of other catalysts and/or solvents, 11 as well as attempts at dissolving metal reduction, 12 gave mainly products resulting from hydrogenolysis of the allylic alcohol function. Separation of these isomers was postponed until after the final ring closure. The alcohols 12a and 12b were converted with N-bromosuccinimide and triphenylphosphine into the bromides 13a and 13b, which were then cyclized to afford norseychellenone 14a and epinorseychellenone 14b. These isomeric ketones could be easily and quantitatively separated by column chromatography on silica gel with the desired isomer eluting first (10% EtOAc/PhH): 14a, R_f 0.35; 14b, R_f 0.25. The isomers were clearly distinguished by their 200-MHz NMR spectra as follows: 14a (CCl₄), δ 1.2-2.4 (m, 13 H), 0.97 $(s, 3 H), 0.94 (s, 3 H), 0.79 (d, 3 H, J = 7 Hz); 14b (CCl₄), \delta$ 1.2-2.4 (m, 13 H), 1.12 (d, 3 H, J = 7 Hz), 1.02 (s, 3 H), 0.98(s, 3 H). The identity of structure 14a was proven by comparison (NMR, IR, TLC) with an authentic sample of norseychellenone.¹³ The synthesis was completed by reaction of 14a with methyllithium followed by dehydration with thionyl chloride according to the procedure of Piers³ to give 1 in quantitative yield, thus ending a 10-step synthesis of seychellene (1) from 2,3-dimethylcyclohexenone (2) in 20% overall

The utility of this synthetic approach to highly functionalized bicyclo[2.2.2] octane derivatives, as demonstrated by this direct total synthesis of seychellene, should be quite high and is currently under investigation in our laboratory.

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References and Notes

- (1) This work was presented at the 13th Western Regional Meeting of the American Chemical Society, Anaheim, Calif., Oct 1977, Abstract 227.
- (2) (a) E. Guenther, "The Essential Oils", Vol. III, Van Nostrand, New York, N.Y., 1949, p 569, (b) N. Tsubuki, K. Nishimura, and Y. Hirose, Bull. Chem. Soc. Jpn., 40, 597 (1967). (c) G. Wolff and G. Ourisson, Tetrahedron, 25, 4903 (1969); Tetrahedron Lett., 3849 (1968).
- E. Piers, R. W. Britton, and W. de Waal, J. Am. Chem. Soc., 93, 5113 (1971);
 Chem. Commun., 1069 (1969).
- (4) K. J. Schmalzl and R. N. Mirrington, Tetrahedron Lett., 3219 (1970); R. N. Mirrington and K. J. Schmalzl, J. Org. Chem. 34, 2358 (1969).
- Mirrington and K. J. Schmalzl, J. Org. Chem., 34, 2358 (1969).
 N. Fukamiya, M. Kato, and A. Yoshikoshi, Chem. Commun., 1120 (1971).
- (6) G. Frater, *Helv. Chim. Acta,* **57**, 172 (1974).
- (7) M. E. Jung and C. A. McCombs, Tetrahedron Lett., 2935 (1976); Org. Synth., 58, in press.
- Several other groups have also made use of silyloxydienes: 8a-c. (a) S. Danishefsky and T. Kitahara, J. Am. Chem. Soc., 96, 7807 (1974); J. Org. Chem., 40, 538 (1975); S. Danishefsky, R. K. Singh, and T. Harayama, J. Am. Chem. Soc., 99, 5810 (1977); S. Danishefsky, P. E. Schuda, T. Kitahara, and S. J. Etheredge, ibid., 99, 6066 (1977), and 98, 3028 (1976); S. Danishefsky, C. F. Yan, and P. M. McCurry, Jr., J. Org. Chem., 42, 1819 (1977). (b) G. M. Rubottom and D. S. Krueger, Tetrahedron Lett., 611 (1977).
 C. T. Ibuka, Y. Mori, and Y. Inubushi, ibid., 3169 (1976).
- (c) T. Ibuka, Y. Mori, and Y. Inubushi, *Ibid.*, 3169 (1976).

 (9) Prepared from 2-methylcyclohexane-1,3-dione by formation of the isobutyl enol ether (*I*-BuOH, TsOH, PhH, \(\Delta\); 88%) followed by addition of methyllithium and acidic hydrolysis (95%). See also D. Nasipuri and M. Guha, *J. Indian Chem. Soc.*, 41, 243 (1964); G. V. Kondrat'eva, G. A. Kogan, T. M. Fadeeva, and S. I. Zav'yalov, *Izv. Akad. Nauk SSSR, Ser. Khim.*, 1648 (1964); J. M. Conia and F. Rouessac, *Bull. Soc. Chim. Fr.*, 1925 (1963).
- (10) We assume that the stereochemistry of the secondary methyl group α to the ketone is anti to the alkyl residue as shown in 7, 8, and 9. This anti stereochemistry would be expected to be the more stable one. Refluxing a solution of 7 with NaOMe in MeOH for 14 h does not change the 200-MHz NMR spectrum of this compound, nor is the 200-MHz NMR spectrum of 9 altered after similar treatment for 5 h.
- (11) Several catalyst–solvent systems were employed to no avail: Pd/C-EtOAc; Pd/C, NaNO₂-EtOAc; Pd/C, NaNO₂-EtOH; PtO₂-EtOAc; Rh/Al₂O₃-EtOAc; Rh/Al₂O₃-EtOH.
- (12) The α,β-unsaturated aldehyde and ester corresponding to 9 (prepared by oxidation of 7 and 9, respectively) each afforded, upon dissolving metal reduction, a complex mixture of products which contained only minor amounts of the alcohols 12a and 12b.

(13) We are very grateful to Professor E. Piers for a generous sample of (土)norseychellenone.

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Nuclear Magnetic Resonance Manifestation of the Configurational Differences between the L- and D-Norepinephrine Complexes with Cobaltous Adenosine 5'-Triphosphate, an Aqueous Chiral Shift Reagent

Sir:

Enantiomers in achiral environments give rise to identical NMR spectra. Spectral resolution of enantiomers has been achieved by employing chiral solvents1 or by applying chiral paramagnetic lanthanide shift reagents.² Hitherto these methods have been restricted to nonaqueous media. ATP (and its metal chelates) contains the chiral D-ribose moiety and upon association with enantiomers should provide the chiral environment necsssary for spectral resolution. However, the chemical shifts induced by ATP in the protons of catecholamines are rather small, 0.3 ppm or less.³ Thus the addition of ATP to the racemic mixture of norepinephrine, (OH)2- $C_6H_3C_\beta HOHC_\alpha H_2NH_3^+$, resulted in measurable shifts for all of the protons similar to those previously obtained with the pure L enantiomer³ but no spectral resolution into enantiomers was observed. Analysis of the chemical-shift data yielded formation constants for association with ATP and intrinsic shifts in the complexed state similar to those for pure L-norepinephrine³ indicating that within the limits of resolution both enantiomers are equally complexed. This observation is in accord with the finding that complexes of catecholamines with ATP are stabilized mainly by ring stacking and by electrostatic interaction between the ammonium and phosphate groups^{3,4} and suggests that these interactions are present in the ATP complexes of both of the norepinephrine enantiomers. It is reasonable to assume, therefore, that the gross structures of the complexes are similar. In view of the structural model proposed for the ATP complexes of catecholamines,³ this similarity obviously implies different dispositions of only the substituents on the β -carbon atom relative to the ATP molecule. Schematic models of the structures of the ATP complexes with L- and D-norepinephrine are shown in Figure 1. On the basis of these models spectral resolution of enantiomers is expected for the β proton. Unfortunately this proton is not only subject to relatively small induced shifts but its resonance is obscured by the residual HDO signal of the solvent.

The chelation of a divalent metal ion by ATP results only in a slight reduction in the complexing ability of the latter for catecholamines and in minor alteration of the structure of the catecholamine-ATP complex, while offering the advantage of large dipolar shifts when Co²⁺ is the cation.⁴⁻⁶ The effect of the cobaltous ATP chelate on the aliphatic portion of the proton NMR spectrum of racemic norepinephrine is shown in Figure 2, where for comparison the spectrum of L-norepinephrine taken under similar conditions is also given. It is seen that two sets of resonances are observed with the racemic mixture: one for each enantiomer. No separation into enantiomeric signals was observed in the aromatic portion of the spectrum. At the lower temperature both the extent of association and the intrinsic shifts are enhanced resulting in higher resolution. Although the induced shift in the β proton is not the largest, it exhibits the highest dispersion as anticipated from

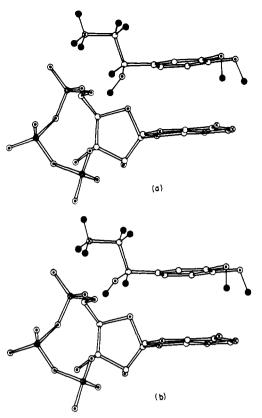


Figure 1. Schematic representation (side view) of the preferred structures of the ATP complexes with (a) L-norepinephrine and (b) D-norepinephrine

Table I. Chemical Shifts a Relative to the Uncomplexed State of the 1:1 Complexes of L- and D-Norepinephrine with CoATP at 27 °C

proton	L-NE	D-NE
H ₂ , ring	2.09	2.09
H ₅ , ring	2.09	2.09
H ₆ , ring	3.05	3.05
	4.40	3.49
$egin{array}{c} H_{oldsymbol{eta}} \ H_{lpha} \end{array}$	4.26	4.66
$H_{\alpha'}$	5.96	5.96

a In parts per million.

the structural models (cf. Figure 1). Since similar concentrations were employed for both samples, the facts that the protons of the L enantiomer experience the same chemical shifts when taken alone or in the racemic mixture (compare traces a and b in Figure 2) and that the aromatic protons remain unseparated indicate that the complex formation constants for the two enantiomers are the same. A formation constant of $15 \pm 3 \,\mathrm{M}^{-1}$ at 27 °C was determined from titrations of fixed norepinephrine concentrations with CoATP in a manner similar to that previously described.⁵ The chemical shifts (upfield relative to the uncomplexed state) of the norepinephrine-CoATP complexes resulting from the data analysis are summarized in Table I. The line width of the β proton in the complexed state is 21 Hz for L-norepinephrine and 13 Hz for the D enantiomer. The former value is in good agreement with the value of 19.8 Hz calculated from the longitudinal relaxation rate,⁵ indicating that the line width is governed by the dipolar interaction with the Co²⁺ ion. A distance of 4.9 Å between the β proton of Lnoreinephrine and the cobaltous ion of CoATP has previously been determined from the relationship

$$1/T_1 = Cr^{-6}f(\tau_c)$$